

This is a protocol for production of dsRNA to be injected in worms for RNAi.

Notes on primers

1. when designing primers, keep in mind that you should amplify a region which is 80% or more unique (at DNA level), in order to preserve specificity;
2. the forward and reverse primers should have the following extensions (19 bases each):
direct primer: 5'-TAATACGACTCACTATAGG-your_seq-3'
reverse primer: 5'-AATTAACCCTCACTAAAGG-your_seq-3'

Steps

1. **PCR** your fragment of interest. Make the following mix:
 - 10x PCR buffer 10 μ l
 - Forward primer (25 μ M) 2 μ l
 - Reverse primer (25 μ M) 2 μ l
 - Template (e.g., 0.5ng genomic) 1 μ l
 - 10mM dNTPs 2 μ l
 - DNA polymerase 1 μ l
 - MQ water 82 μ l
2. PCR conditions
 - 92°C 1min
 - 92°C 20 sec
 - anneal 40 sec
 - extend XX sec
 - 72°C 7 min
 - 4°C hold
3. Annealing temp. depends on primer; extension temp. and time depend on the polymerase used. Run 2 μ l of each reaction on an agarose gel. Clean the PCR reactions using the Qiagen PCR cleanup kit; elute with 50 μ l.
4. **T3 and T7 transcription reactions**, using the Ambion kit Mix in the order below, at room temperature:
 - NTPs 10 μ l
 - 10X T3 OR T7 buffer 2.5 μ l
 - DNA template 10 μ l
 - T3 OR T7 enzyme mix 2.5 μ l
5. Incubate at 37 °C for 3-5 hours
6. **Remove template DNA**: add 1.3 μ l DNase to each reaction, incubate at 37°C for 15 minutes
7. Take 2 μ l samples of each reaction, mix with 2 μ l of sample buffer and 2 μ l or RNase-free H₂O
8. **Clean the reactions** using the RNeasy kit. Elute each reaction with 30 μ l RNase-free water into the same tube. [alternatively, you could mix the two reactions and clean both in one round; this spares columns and, especially, pipetting work]
9. **Anneal the RNAs**: Add 30 μ l of 3X injection buffer to each tube. Incubate at 68°C for 10min and then at 37°C for 30min. Take 2 μ l of each sample for a gel and treat it as above.
10. **Check the RNAs**: Dilute 50x an aliquot of the final dsRNA solution and measure the OD₂₆₀. (Assume that a solution of 50 μ g/ml gives an OD=1). Final yield should be around 1..2mg/ml. Run an agarose gel with the three samples per reaction. If everything is ok, proceed to injection.

Solutions

3X injection buffer

1. 120mM KPi pH 7.5
2. 18mM K citrate pH 7.5
- 4% PEG 6K, filter 0.2 μ m